Phytochemical Composition and *In vitro* Anti-Proliferative Activity of *Oxygonum sinuatum* (Meisn.) Dammer on Selected Cancerous Cells

Douglas Kahura Njuguna¹, Karori Mbuthia¹, Chrispus Mutuku², Mercy Jepkorir³, Jecinta Wanjiru Ndung'u⁴, Reginah Mwangangi⁵, Jean Chepngetich⁶ and Peter Mwitari²*

¹Department of Biochemistry and Molecular Biology, Egerton University, P.O.Box 536, Egerton, Kenya.
²Center for Traditional Medicine and Drug Research, KEMRI, P.O.Box 54840-00200, Nairobi, Kenya.
³Department of Chemistry and Biochemistry, Lakipia University, P.O.Box 1100, Nyahururu, Kenya.
⁴Department of Biochemistry, Rongo University, P.O.Box 103-40404, Rongo, Kenya.
⁵Department of Biochemistry and Biotechnology, Kenyatta University, P.O.Box 43844-00100, Nairobi, Kenya.
⁶Department of Biochemistry, Jomo Kenyatta University of Agriculture and Technology, P.O.Box 6200-00200, Nairobi, Kenya.

Authors’ contributions

This work was carried out in collaboration between all authors. Authors DKN, CM and JC designed the study and wrote the protocol. Authors DKN, PM and JC performed the statistical analysis and wrote the first draft of the manuscript. Authors KM and PM managed the analyses of the study. Authors MJ, JWN and RM managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JOCAMR/2018/44413

Editors:
(1) Dr. Aditi Singh, Amity Institute of Biotechnology, Amity University, Lucknow, India.

Reviewers:
(1) Ahmed Mohammed Abu-Dief Mohammed, Sohag University, Egypt.
(2) Daohong Chen, China.
(3) Ramaiah Maddi, Hindu College of Pharmacy, India.

Complete Peer review History: [http://www.sciencedomain.org/review-history/26695](http://www.sciencedomain.org/review-history/26695)

Received 07 July 2018
Accepted 30 September 2018
Published 19 October 2018

*Corresponding author: E-mail: pmwitari@kemri.org;*
ABSTRACT

Cancer is a leading cause of deaths worldwide. The current search for alternative drugs is imperative. Medicinal plants are the most promising source of new anticancer agents. This study evaluated the phytochemical composition, and anti-proliferative activity of Methanol-Dichloromethane (1:1) extracts from the leaves, stem and fruits of *Oxygonum sinuatum*. Phytochemicals screening was done using standard procedures which were based on color change and/or precipitation. The MTT assay was used to test for the anti-proliferative activity of the plant extracts on selected cancer cell lines. In addition, normal Vero cells were used. 5-flourouracil was used as the standard drug. Results from this study revealed that stem, leaves and fruits extracts contained alkaloids, glycosides, saponins, tannins, flavonoids, terpenoids, steroids and phenols. The leaves, stem and fruits extracts showed moderate anti-proliferative activity on 4T1, Hcc 1395, 22Rv1 and DU145 with the best activity been registered by fruits extract on 4T1 cell line; 35.84±3.549µg/ml. The fruit extract showed no activity on DU145 cell line. The leaves and fruit extract showed no toxicity to Vero normal cells (CC50 >1000µg/ml) however, the stem extract was toxic to Vero cell, CC50 413.73±21.022 µg/ml. The highest selectivity index was registered by fruit extract on 4T1; SI=27.90. The leaves and stem extracts also showed high selectivity (SI >3) on 22Rv1 and DU145 respectively. The anti-proliferative activity of *Oxygonum sinuatum* on the selected cancer cell lines can be attributed to the phytochemical compounds present in the plant extracts.

Keywords: *Oxygonum sinuatum*; anti-proliferation; phytochemicals; selectivity index (SI).

1. INTRODUCTION

Cancer is one of the leading causes of deaths worldwide accounting for 8.2 million deaths annually, which is approximately, 13% of the total deaths [1]. New cancer cases per year are estimated to be 14 million a figure predicted to rise to 22 million within the next two decades [1]. Regrettably, 70% of the total cancer deaths occur in low and middle income countries [2]. The major cancer predisposing factors include; tobacco use, excessive alcoholism, some infectious agents, obesity, physical inactivity and westernized diet [3]. In Africa, cancer mortality rate is greater than that of HIV, TB, and malaria combined [4]. Cancer causes 7% of the total national mortality annually in Kenya ranking third after infectious and cardiovascular diseases [5]. The poor prognosis of cancer is due to lack of enough facilities for diagnosis and treatment, limited access to health facilities and high cost of conventional treatment [6]. The major conventional treatment methods include chemotherapy, radiotherapy, immunotherapy and surgery. Despite being expensive and beyond reach by majority of people in poor countries, these methods are also laden with various side effects [1]. This has necessitated for the search of alternative cancer management and treatment agents.

In Africa, up to 80% of the population relies on the use of herbal medicine for their primary healthcare [7,8]. The conventional medicine in Kenya provides for only 30% of the population, this means that more than two-thirds of Kenyans rely on traditional medicine for their healthcare needs [9]. Many people perceive medicinal plants to be safer than their synthetic alternatives. Wide range of plant extracts have been used as raw or processed drugs in the treatment of various diseases. The different parts used include root, stem, flower, twigs, exudates as well as whole plant [10]. Various phytochemical compounds found in medicinal plants have been a source of innumerable therapeutic agents and a tool for development of conventional medicine [11].

*Oxygonum sinuatum* (Meisn.) Dammer (Polygonaceae) is an annual weed growing in the drier areas and widely distributed in Kenya [12]. *O. sinuatum* is a decumbent plant, its stem grows up to 90 cm long [13]. The plant is polygamous with prickly fruits. Traditionally the plant is used in treatment of various diseases including cancer [14-17]. However, its anti-proliferative potential has not been fully investigated. Previous studies on methanolic extract of the whole plant demonstrated antioxidant and anti-inflammatory activity as well as broad spectrum antibacterial effect [18]. A different study by Uma and Bharti [19] correlated the antioxidant activity of methanolic extract from the whole plant to phenolic compounds in the plant. Kamuhabwa et al. [20] showed the anti-proliferative potency of the whole plant methanolic extract on skin and colon carcinomas.
2. MATERIALS AND METHODS

2.1 Plant Collection

The plant was collected from Ngo’ng Hills (1.3618°S, 36.6566°E) Kajiado County and transported to the Center for Traditional Medicine and Drug Research CTMDR, KEMRI for taxonomic identification and processing. A botanical sample was deposited at CTMDR, KEMRI and given the voucher number KEMRI/CTMDR/DKN/1/2016. The plant parts were sorted into leaves, stems, and fruits, dried at room temperature and ground into fine powder using a laboratory mill.

2.2 Extraction Procedures

The grounded samples were weighed using a top balance and put in a 500 ml flat bottomed flask. Methanol and dichloromethane (DCM) (1:1) was added until the plant material was completely submerged. The mixture was then agitated for thorough mixing then left to extract for 24 hours with frequent shaking to ensure effective extraction. After the 24 hours the mixture was filtered using Butchner funnel; Whatman no. 1 filter paper. The extracts were then concentrated using a rotary vacuum evaporator with a water bath at 40°C. The concentrated extracts were stored at 4°C until use.

2.3 Qualitative Phytochemical Analysis

The extracts’ phytochemicals composition was tested using standard procedures with slight modifications of the procedures [21,22].

Alkaloids; Mayer’s Test: About 2 ml of the plant extract was put in a clean test tube. Few drops of Mayer’s reagent were added and observations made. Appearance of white precipitate indicates presence of alkaloids.

Cardiac glycosides; Keller Killiani Test: Approximately 2 ml of the plant extract was put in a test tube, 1 ml of glacial acetic acid containing 1 drop of ferric chloride was added. This mixture was added to 1 ml of concentrated sulphuric acid and observations made. Formation of a brown ring indicates presence of cardiac glycosides.

Saponins: The crude extract was mixed with 5 ml of water and vigorously shaken. The mixture was left to stand for about 15 seconds. Formation of a stable form indicates presence of Saponins.

Tannins: To about 0.5 g of the plant extract in a test tube, 20 ml of double distilled water was added and heated to boiling. The mixture was then filtered and 1% FeCl3 was added to the filtrate and observation made. A brownish green coloration indicates the presence of tannins.

Flavonoids: To a portion of the plant extract in a test tube, 5 ml of dilute ammonia was added followed by addition of 2 ml of concentrated sulphuric acid. The appearance of a yellow color indicates the presence of flavonoids.

Terpenoids: About 2 ml of the plant extract was put in a test tube. To this, 2 ml of chloroform was added followed by 1ml of concentrated sulphuric acid. Observations were then made. A brown ring at the junction of two layers indicates the presence of terpenoids.

Steroids; Liebermann Rurchard Reaction: To a sample of the plant extract in a test tube, 10 ml of chloroform was added and filtered. To 2 ml of the filtrate, 2 ml of acetic acid was added followed by addition of concentrated sulphuric acid. The formation of blue-green ring at the junction of two layers indicates presence of steroids.

Phenols: Portion of the plant extract was put in a test tube and treated with few drops of 2% ferric chloride. Observations were then made. The appearance of blue green coloration indicates presence of phenols.

2.4 Anti-proliferative Assay

2.4.1 Sample preparation

Briefly, 10 mg of the plant extracts (leaves, stem, and fruits extracts) were weighed in a 1.5 ml eppendorf tube using analytical balance. To each sample 100 µl dimethyl sulfoxide (DMSO) solution was added and the mixture vortexed, 900 µl of PBS was added to the mixture to make 1 ml of the solution.

2.4.2 Cell line culturing

Vero cell line (normal) and 4T1 (breast), 22Rv1 (prostate), Hcc 1395 (prostate), DU 145 (breast) cancer cell lines (ATCC) were obtained from Center for Virology Research (CVR), KEMRI. The cells were revived in a water bath at 37°C and cultured in T-75 flasks with Growth Essential Medium (GEM, SIGMA USA) supplemented with 10% Fetal Bovine Serum (FBS) and 100 µg/ml
streptomycin then incubated for 72 hours to attain confluence. All incubation was done at 5% CO₂ and 37°C [23].

2.4.3 MTT assay

Upon attainment of confluence, both Vero and cancer cells were washed using Phosphate Buffer Saline (PBS) and harvested by tripsinization. The number of viable cells was determined by Trypan blue exclusion test. Approximately 2.0 ×10⁶ cells/ml suspension for both the Vero and cancer cells were seeded in 96-well plates and incubated for 24 hours for the cells to attach to the plates. Briefly, 15 µl of the test sample extracts at seven different concentrations each serially diluted was added from rows H to B and the plates incubated for 48 hours. Row A acted as the negative control. The viability of the cells after extracts addition and incubation was determined using MTT cell proliferation assay which is based on the ability of the living cells to reduce the yellow 3-(4,5-dimethyl-2-thiazoly)-2, 5-diphenyltetrazolium bromide (MTT) dye to a purple formazan product [24,25,26]. After 48 hrs 10 µl MTT dye was added to the cells and incubated for 4 hrs. All the media was then removed from the plates and 50 µl of DMSO was added to solubilize the formazan product. Absorbance was read on a scanning multi well spectrophotometer at 562 nm [27]. The inhibitory concentration (IC₅₀) on cancer cells and cytotoxic concentration (CC₅₀) on normal Vero cells was determined [28]. The Selectivity index (SI=CC₅₀/IC₅₀) was also calculated.

2.5 Data Analysis and Presentation

Analysis of statistical data obtained from this study was done using Cruzi 7 Drug Cytotoxicity Software and Statistical Package of Social Science (SPSS Version 20, IBM, USA). The difference between the treatment and the control in this study was tested for statistical significance using one-way analysis of variance. A value of p≤ 0.05 was considered significant. The IC₅₀ and CC₅₀ values were expressed as Mean ± Standard deviation (SD). Tables were used to enhance the presentation of the results.

3. RESULTS

3.1 The Yield of the Extracts

The plant extracts yield was calculated and recorded as a percentage. The fruit extract had the highest yield of 11%, the leaves extract had 10% while the stem extract had the lowest yield of 5%.

3.2 The Phytochemistry Results

Phytochemical analysis revealed the presence of alkaloids, glycosides, saponins, tannins, flavonoids, terpenoids, steroids and phenols in all the extracts. The tests were based on precipitation and color change. Table 1 illustrates these results.

3.3 The Anti-proliferative Activity

The three plant parts extracts registered significantly higher anti-proliferative activity (P≤0.05) on 4T1, 22Rv1, Hcc 1395 and DU145. The highest activity was registered by fruit extract on 4T1 with an IC₅₀ of 35.84±3.549 µg/ml which was statistically comparable to that of the reference drug on the same cell line. The fruits extract had no activity on DU145 cell. The stem extract was toxic to Vero cells while the leaves and fruit extract showed no toxicity to the Vero cells (CC₅₀ >1000 µg/ml). These results are illustrated in Table 2.

4. DISCUSSION

The leaves, stem and fruits extracts of O. sinuatum showed anti-proliferative effects against 4T1 (mouse breast cancer cell), Hcc 1395 (human breast cancer cell), 22Rv1 (human prostate cancer cell) and DU 145 (human metastatic prostate cancer cell) as depicted by the IC50 values. An IC50 value is the amount of the concentrated extract required to inhibit 50% of the cell growth. The stem extract showed toxicity to Vero cell line, this could be attributed to the presence of other compound(s) in the stem extract which were not investigated in this study. According to the American National Cancer Institute (NCI) drug screening program, a crude extract is considered to have a better in vitro anti-proliferative activity if the IC₅₀ value is less than or equal to 30µg/ml following incubation for 48 hours to 72 hours. A crude extract with an IC₅₀ of between30µg/ml to 1000 µg/ml is considered to have moderate activity [29,30,31]. Therefore, the three extracts (leaves, stem and fruit extracts) had moderate activity on the four cancer cell lines used in this study as illustrated in Table 1. The extracts IC₅₀ mean values were statistically different to the IC₅₀ mean values of the 5-flourouracil - a standard anticancer drug except for the fruit extract on 4T1 (breast cancer cell line) whose IC₅₀ was statistically comparable to that of 5-flourouracil on the same cell line.
Table 1. Phytochemical screening results and observations

<table>
<thead>
<tr>
<th>Phytochemical tested</th>
<th>Leaves</th>
<th>Stem</th>
<th>Fruits</th>
<th>Observations</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkaloids</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>White precipitate</td>
</tr>
<tr>
<td>Glycosides</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Formation of a brown ring</td>
</tr>
<tr>
<td>Saponins</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>A stable foam</td>
</tr>
<tr>
<td>Tannins</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Brown-green colouration</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Yellow colouration</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Brown ring at junction of 2 layers</td>
</tr>
<tr>
<td>Steroids</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Blue-green ring at junction of two layers</td>
</tr>
<tr>
<td>Phenols</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Blue-green colouration</td>
</tr>
</tbody>
</table>

Key: + indicates the presence of the phytochemical tested

Table 2. Anti-proliferative assay results; CC50s, IC50s, and selectivity indices (SI) of the methanol-dichloromethane leaves, stem and fruits extracts from O. sinuatum on Vero (normal), 4T1, 22Rv1, Hcc 1395 and DU 145 cancer cells

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Plant part</th>
<th>Reference drug</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vero (Normal)</td>
<td></td>
<td>5-Flourouracil</td>
</tr>
<tr>
<td>CC50 (µg/ml)</td>
<td>&gt;1000</td>
<td>413.733±21.022a</td>
</tr>
<tr>
<td>IC50 (µg/ml)</td>
<td></td>
<td>222.792±2.220a</td>
</tr>
<tr>
<td>SI (CC50/ IC50)</td>
<td>1.341±0.023a</td>
<td>1.857±0.045a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>27.901±1.714a</td>
</tr>
<tr>
<td>4T1 (breast)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IC50 (µg/ml)</td>
<td>745.460±17.126a</td>
<td>222.792±2.220a</td>
</tr>
<tr>
<td>SI (CC50/ IC50)</td>
<td>35.841±3.549a</td>
<td>745.460±17.126a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1.734±0.023a</td>
</tr>
<tr>
<td>22Rv1 (prostate)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IC50 (µg/ml)</td>
<td>181.480±13.972a</td>
<td>956.966±10.996a</td>
</tr>
<tr>
<td>SI (CC50/ IC50)</td>
<td>5.543±0.427a</td>
<td>1.045±0.012a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1.341±0.023a</td>
</tr>
<tr>
<td>Hcc 1395 (breast)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IC50 (µg/ml)</td>
<td>867.058±7.354a</td>
<td>529.199±12.707a</td>
</tr>
<tr>
<td>SI (CC50/ IC50)</td>
<td>114.870±15.825a</td>
<td>367.487±16.108a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>1.045±0.012a</td>
</tr>
<tr>
<td>DU 145 (prostate)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IC50 (µg/ml)</td>
<td>559.292±16.532a</td>
<td>114.870±15.825a</td>
</tr>
<tr>
<td>SI (CC50/ IC50)</td>
<td>1.341±0.023a</td>
<td>1.045±0.012a</td>
</tr>
</tbody>
</table>

Key: (superscript) indicates that the IC50 mean values are statistically different compared to those of the reference drug:

This study is consistent with previous studies by Kamuhabwa et al. [20] which showed that the whole plant methanolic extract of O. sinuatum had anti-proliferative activity against HT29 (colon adenocarcinoma) and A431 (skin carcinoma) but registered no activity on HeLa (Cervical cancer cells). Another study by Crawford et al. [33] showed that the crude methanolic extract from O. sinuatum inhibited mammalian endothelial cell proliferation and migration. The anti-proliferative activity of the three plant extracts against the four cancer cell lines was not directly correlated to the phytochemical profile of the plant. However, this activity can be attributed to the presence of

The selectivity index (SI) of an extract shows its ability to inhibit the growth of cancer cells more than it does to the normal (Vero) cell line. The (SI) of an extract is calculated as a ratio of the CC50 on the Vero cell line to the IC50 of the cancer cell line. An extract with the SI greater than three is considered to be highly selective [32] and may indicate the potential use of the plant extract in cancer management. The highest selectivity index was registered by fruit extract on 4T1 (SI=27.90). The leaves and stem extracts also showed high selectivity (SI >3) on 22Rv1 and DU145. The extracts selectivity indices ranged from 1.04 to 27.90.
various secondary metabolites that were found to be present in the three plant extracts as shown in Table 1. In addition to this, other studies have shown that *O. sinuatum* contains anthraquinones, diterpenoids and phenols [19,33]. Various phytochemical compounds have been isolated and shown to have anti-proliferative activity against various cancer cell lines [34,35].

Alkaloids are a group of cyclic compounds that have a nitrogen atom in their chemical ring structure. Alkaloids have been isolated from various plants and their therapeutic activity tested. Vinblastine and vincristine are alkaloids isolated from the Madagascar periwinkle *Cantharanthus roseus*. They were the first alkaloids to be used as anticancer agents [34]. Cardiac glycosides are effective therapies for congestive heart failure and arrhythmia [36]. Recently, cardiac glycosides have been documented to have anti-proliferative activity on human carcinoma and leukemia cells [37]. Saponins have also been documented to have anti-proliferative effect on various cancer cells [38].

Phenolic compounds form a large group of molecules including the flavonoids, tannins and coumarins [39]. Phenolic compounds are known for their antioxidant and anti-proliferative activity on various cancers including leukemia, colon adenocarcinoma, cervical carcinoma prostate and bladder carcinomas [39]. Phenols from whole plant methanolic and aqueous extracts of *O. sinuatum* showed a high antioxidant activity [19]. Flavonoids are a group of water soluble compounds containing 15 carbon atoms. Takemura [40] provided a comprehensive review of chemoprevention by flavonoids on breast cancer. Flavonoids isolated from flowers of Tecoma stains demonstrated significant antitumor activity on Hep-2 (Human larynx) carcinoma cell line [41]. Tannins a subclass of hydroxybenzoic acids also manifests anti-proliferative activity against human hepatocellular carcinoma, human prostate cancer, human cervical cancer and mouse sarcoma cell lines [42]. Terpenoids and steroids are unsaturated cyclic or linear hydrocarbons with a varying number of isoprene units. Corticosteroids have been in use in management of cancer [43]. A Study by Joao [44] showed that oxysterols were active against LAMA-84 (leukemia) cells.

Although this study did not directly depict the exact mechanisms of action of the plant extracts, Crawford et al. [33] showed that crude methanolic extract of *O. sinuatum* have anti-angiogenic activity both in vitro and in vivo. Emodin, an anthraquinone and coleon A lactone, a diterpenoid from *O. sinuatum* [33] has been shown to be an inhibitor of protein tyrosine kinase [45], casein kinase II [46] and human epidermal growth factor receptor 2 (HER-2) [47] all of which are involved in cell division. *O. sinuatum* could have imparted its anti-proliferative activity through the same mechanisms.

5. CONCLUSION

This study provides the phytochemical profile of methanol-dichloromethane stem, leaves and fruits extracts of *O. sinuatum*, and further gives the anti-proliferative activity of the three plant extracts against breast and prostate cancer cell lines. The anti-proliferative activity of these extracts can be attributed to the phytochemicals that are present in the extracts. This study provides important basis for further investigation on the development of herbal medicine from *O. sinuatum* as alternative therapy for cancer treatment and management. This study recommends further studies to isolate the bioactive components from the three extracts and elucidate their mechanisms of action.

CONSENT

It is not applicable.

ETHICAL APPROVAL

Ethical approval was sought from Kenya Medical Research Institute (KEMRI) Centre for Traditional Medicine and Drug Research (CTMDR) Centre Scientific Committee (CSC) and Scientific and Ethics Review Unit (SERU) (KEMRI/SERU/CTMDR/030/3331) before conducting the study.

ACKNOWLEDGEMENT

The authors are very grateful to the Center for Traditional Medicine and Drug Research (CTMDR), KEMRI for their support during the study.

COMPETING INTERESTS

Authors have declared that no competing interests exist.
REFERENCES

   PMID: 18626751
   PMID: 22752881
   (Accessed 20th May 2016)


45. Jayasuriya H, Koonchanok NM, Geahlen RL, McLaughlin JL, Chang CJ. Emodin,


© 2018 Njuguna et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sciencedomain.org/review-history/26695